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Fungal contamination and exposures in different water resources, hazards and remediation

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The fungi population in water has gained attention during the three decades. In this review, article attempt has been made to show its vulnerability as numbers of fungal species are now treated as water contaminants to indicate diseases in the human body. However, the importance of study of fungi has been less focused on surface water, groundwater systems and in agricultural wetlands. Time has come to correlate the population of fungi species isolated and identified from different sources of water-related with the climate change. The number of the fungal species are isolated and identified by the researchers may or may not have common occurrence but those definitely contaminate surface water at the different sources as well as different countries of the globe. In some cases, study has been made to analyze the occurrence of yeasts and filamentous fungi in the water body to investigate their correlation with bacterial indicator and faecal pollution¹.

Keywords: Ground water, climate change, filamentous fungi, faecal pollution, effects on human beings, remediation.

Introduction

Water is a primary requisite for life on earth but contaminants arising from incessant inflow and pollution of water bodies do have a myriad effect on ecology as well as on human body. It is necessary to protect consumers from water-borne diseases. The major problem is to get pathogenfree drinking water. Water utilities, however, rely solely on monitoring indicator bacteria, such as coliform and Escherichia coli (E. coli) to make sure microbiological quality of drinking water. In nature, there are abundance of coliform bacteria but these may not cause diseases. Their existence is often useful to indicate the potentially harmful bacteria may be present in the water whereas faecal coliform and E. coli originate exclusively from human and animal faecal wastes². In this concern, the study has found out over 70,000 fungal species are existing in the environment in which unicellular yeast, filamentous fungi and multi cellular mould which is more or less 300 have been counted may spread the human diseases, and about dozen cause 90% fungal infections. Those involved various types of diseases, including allergies to fungal production of toxins, antigens and some directly invest host. Life-threatening disseminated infectious to other form of mucosal infections are being caused by several species of fungi.

Nowadays, there are reports in the media and the literature regarding occurrence of bacteria, yeast and filamentous fungi in treated and bottled mineral water too. Although causal relationships between fungal occurrence and water quality remains uncertain, a few cases of report being available where it is found fungal bioflims with in the municipal water distribution system. Filamentous fungi like *Penicilium* sp., *Aspergillus* sp. and *Candida* sp. have been investigated from potable water. Arvanitidou *et al.*, 1999 had investigated and isolated filamentous fungi from 82.5% sample and yeast from 11.1% from potable water of hospital and community samples³. It is witnessed an increase awareness about the potential for fungi to degrade complex natural substances due to broad enzymatic capabilities⁴. It was reported of their possible pathogenicity towards susceptible humans, animals. A few fungi produce mycotoxins that can display overlapping toxicities in vertebrates, plants and microorganisms⁵.

A study shows, total and faecal coliforms, and total heterotrophic bacterial counts were estimated in parallel with counting of filamentous fungi and yeast in groundwater, in addition to some physicochemical parameters⁶.

It is noticeable that total faecal coliform and heterotrophic bacteria estimated in a parallel method while studying or counting of filamentous fungi and yeast in bottled water and municipality's tap water⁴.

It is interesting to note that developing countries are not taken into consideration of vulnerability of yeast and faecal coliforms, the measures and effects of mycotoxins in the human body, and more importantly resources of toxicological data, the analytical capacity to enforce regulations are less studied^{7,8}.

Discussion

In this review article, it has been observed that groundwater, surface water and other resources of water have contaminants of bacteria, filamentous fungi and other microorganisms which cause health hazard to human being and climate change may cause pollution of water higher in the last three decades.

Usually, microbial quality of water is measured by the microorganism using indicator mainly *Escherichia coli*⁹. However, bacterial indications are importantly determined for sanitary and public health safety, are more challenging while enterovirus and protozoa are more resistant against disinfection, evenmore, *E. coli* zero count doesn't signify the absence of other microorganisms^{10,11}. Hence, heterotrophic plate counting method is a prime procedure for measuring the presence of fungi as it is used to indicate the concentration of changes of microorganism whether it is entering or growing in the treated drinking water or not^{12,13}.

Based on this above prospects, it is worth mentioning that many countries till are not taken into consideration the importance of fungi infection in treated drinking water. Further to that, the United State Environmental Protection Agency created the purpose of protecting human health and the environment by planning and implementing regulation law passed by the Congress¹⁴. Even more, EU Drinking Water Regulations (2004) implemented few terms and conditions for maintaining the standard of drinking water but the mentioned authorities did not concern about the treatment of fungi in water, while Sweden is the only country that currently measuring the fungi contaminations in treated drinking water. The Swedish Water regulation authorities set a permissible limit of treated drinking water that is 100 CFU of microfungi in 100 ml water is considerable and safe for human consumption¹⁵.

Growth of fungi based on different sources and physico-chemical parameters

The colony of fungi, their growth, and bioremediation process are dependent on the pH value of water. It has been observed that the positive correlation between aquatic hyphomycetes and the value of pH which ranges in between 5 to 7 and confirmed cases found in groundwater studies while lower pH means acidic water have an influence for binding of heavy metals like manganese and cadmium to the fungal cell wall components, which is a boon for some fungi¹⁶.

The amount of organic substance in water is related to the sources and the surface area of the water bodies. Small surface of water receives the most organic matter due to plant vegetation. Furthermore, surface water with a slow stream contains a rich amount of nitrate, nitrite, phosphate and other products of organic material degradation, such as debris, lignin, hemicelluloses, and cellulose^{17,18}. Contrarily, human activates the water pollution along with fertilizers or industrial and house hold waste^{19,20}. Even more, surface water contains high biomass and rich diversity of plant is degraded by filamentous fungi²¹. Hence survey has been conducted Globally and many European territories in order to get the presence of isolated fungi species in surface, ground, tap water, drinking water, municipal water bodies, and in water distribution systemetc belong to ascomycetous genera Alternia, Aspergillious, Cladosporium, Fusarium and Gibberrella, etc. (Table 1)^{16,22}.

Therefore, the correlation results between filamentous

| | | different water resource | | |
|----------|------|--------------------------|----|--|
| . | | | -, | |

| | | Table 1. Source and | isolated fungi | |
|------------------------------------|-----------------------|--|---|------|
| Country | Year | Sources | Isolated fungi | Ref. |
| Greece, Thessaloniki | 1998 | Tap water (hospital zone) | Alternia sp., Aspergillius sp., Exophilia sp., Trichoderma sp | . 3 |
| UK and USA | 1996 | Surface water | Alternia sp., Aspergillius sp., Mucors sp., Phialophora sp., Pythium sp. | 49 |
| Greece | 2000 | Municipal water | Pennicillium sp., Aspergillius sp., Verticillium sp., Actinomycetales sp., Alternia sp., Cryptococus sp., Curvularia sp., Daratomyces sp., Helminthosporium sp. | 23 |
| Poland | 2000-2002 | Water Distribution System | Aspergillius sp., Cladosporium sp., Fusarium sp., Phialopphora sp., Sesquicikkium sp. | 50 |
| Germany, North Rhine-Westphalia | 1998/9 (12 months) | Drinking water | Phialophora sp., Acremonium sp., Aspergillius sp., Cladosporium sp., Chalara sp. | 48 |
| Portugal | 2003-2004 | Tap water | Acremonium sp., Alternia sp., Aspergillius sp. | 51 |
| Pakistan | 30 samples taken | Municipal Water Body | Aspergillus sp., Monodictys sp. | 22 |
| Australia | 2007-2008 | Municipal water | Cladosporium sp., Penicillium sp., Aspergillius sp., Alternaria sp. | 15 |
| Saudi Arab | Once | Treated watersource in hospital and residential houses | Alternia sp., Aspergillius sp., Acremonium sp., Cladosporium sp., Fusarium sp. | 52 |

fungi and total yeasts, and bacterial indicators of pollution (total coliforms, fecal coliforms and fecal streptococci) of studied groundwater samples. The obtained results represented that, filamentous fungi showed a strong proportional correlation (0.727 and 0.885)) with total coliforms, and fecal coli form saverage counts, accordingly, while filamentous fungi showed weak correlation (0.181) with faecal steptoccocci. Furthermore, total yeasts showed weak proportional correlation (0.065) with total coliforms average counts and weak reverse correlation (-0.154 and -0.128) with faecal coliforms and streptococci respectively, while Table 8 shows the correlation results between filamentous fungi and total yeasts

| $\textbf{Table 2.} Sampling sites of groundwater wells in Giza \ Governorate^{6}$ | | | | | | | | |
|---|-----------------|--------------|-------------|--|--|--|--|--|
| Sample Sites | Well names | Sample sites | Well name | | | | | |
| 1 | Mazghouna | 11 | Meat AlKaed | | | | | |
| 2 | Abu Rakhowan | 12 | Nekla | | | | | |
| 3 | AL Badrashin | 13 | Alreka | | | | | |
| 4 | Abuel Nomers | 14 | Gerza | | | | | |
| 5 | Tami | 15 | Kafr Ammar | | | | | |
| 6 | Ezbetsherif | 16 | Al Maktafia | | | | | |
| 7 | Kafr Hamido | 17 | Kafr Hegazy | | | | | |
| 8 | Berush | 18 | Al Rahwey | | | | | |
| 9 | El Sheikh Zayed | 19 | Al Kata | | | | | |
| 10 | Al Manaia | 20 | Galatma | | | | | |

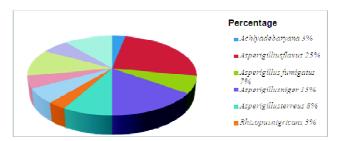


Fig. 1. Relative diversity of filamentous fungal species in groundwater samples⁶.

counts. From the obtained statistical analysis, it can be observed that filamentous fungi showed reverse correlation with all measured physico-chemical factors except temperature and ammonia was proportional. In addition to this, total yeasts showed reverse correlation with some measured physico-chemical parameters (TDS, chloride, temperature, EC, nitrate, total hardness, and iron) and showed proportional correlation with BOD, COD, ammonia, sulphate, turbidity, and pH^6 .

On the other hand, in the recent years in Giza, Egypt, individuals underlying more chronic diseases and immune suppression are exposed to potential pathogens like filamentous fungi and yeast cause the risk of gastroenteritis, pulmonary problems, skin ailments, etc. Now, mostly filamentous

| Giza Governorate ⁶ | | | | | | | | |
|-------------------------------|---------------------|---|--|--|--|--|--|--|
| Parameter | Unit | Measurement method | | | | | | |
| Electric | μs/cm | Conductivity method (APHA, | | | | | | |
| conductivity (EC) | | 2005) using EC meter, Jenway, model: 470 | | | | | | |
| рН | | pH meter (WTW, Model pH, 315i) | | | | | | |
| Temperature | °C | Mercury thermometer, GH Zeal Ltd., London, England | | | | | | |
| Turbidity | NTU | Turbiditimeter [10 b] a potable Hannaturbidimeter (model: HI 93703) | | | | | | |
| Iron | mg/L | Phenanthroline method | | | | | | |
| Ammonia and nitrate | mg/L | A colorimeter (Jenway 6510,En- gland) at 410 nm | | | | | | |
| Chlorides | mg/L | Silver nitrate titrimetric method | | | | | | |
| Sulphate | mg/L | Turbiditimetric method using UV/ Vis spectrophotometer, Unicam model UV4-200 (UK): at wave length 420 mm | | | | | | |
| Total hardness | mg/L | EDTA titrimetric method (APHA, | | | | | | |
| (CaCO ₃) | | 2005) | | | | | | |
| TDS | mg/L | APHA, 2005, Evaporation test method | | | | | | |
| COD | mg/L | Titrimetric method (APHA, 2005) using spectrophotometer (Dr/ 20000) for use at 600 nm | | | | | | |
| BOD | mgO ₂ /L | Winkler's iodometric method (APHA, 2005) | | | | | | |

Table 3. Used parameters and methods in groundwater wells in

fungi and yeast are also considered potentially pathogens and their quantification is practically identical for determining the total counts 23 .

Twenty four species of fungi were isolated from the collection of 40 water samples from Vembanadu wetland agroeco system. The fungi Aspergillius sp. almost significant to mention, was five species, followed by Curvulariasp. and Penicillium sp. (3 species each). The prevalence of Aspergillius niger was noticed (50%) followed by Penicillium sp. (40%), while Paecilomyces sp. and Aspergillius flabus were accounted for 22.5% each accordingly. Hence, detail colonial count of fungi tested (n = 40) in four different consecutive guarter were given in Table 9 and Table 10²⁴.

Public health implication of mycotoxigenic fungi

The presence of fungi are not taken into consideration until the report of fungal contaminated water was noticed in Finland and Sweden during the 1980s and 1990s^{25,26}. Water borne filamentous fungi are acted as a pathogenic and allergenic which have an adverse effect on human health, mostly attacked immune-compromised patients²⁷. Pathogenic fungi are also causing hostile infections which led to high mortality rates^{3,28–31}. Consequently, mycotoxins are fungal poisons as secondary metabolites by the mycelia struc-

| Table 4. Physico-chemical parameters of groundwater samples at Giza, Egypt Governorate ⁶ | | | | | | | | | | | | | |
|---|---------|---------|----------|--------|--------|-----------|----------|-----|------|------|-------|--------|-----------|
| Parameter | Ammonia | Nitrate | Sulphate | Iron | TDS | Chlorides | Total | BOD | COD | pН | Temp. | EC | Turbidity |
| Sample | (Mg/L) | (Mg/L) | (Mg/L) | (Mg/L) | (Mg/L) | (Mg/L) | hardness | | | | (°C) | (s/cm) | (NTU) |
| No. | | | | | | | (Mg/L) | | | | | | |
| 1 | 1.4 | 5.75 | 30.5 | 554 | 435.6 | 108 | 300 | 0 | 8 | 7.36 | 28 | 660 | 0 |
| 2 | 1.86 | 8.27 | 35.6 | 0.67 | 374.4 | 84 | 280 | 0 | 12 | 7.3 | 22 | 567 | 4.64 |
| 3 | 1.42 | 3.42 | 31.1 | 0.67 | 335.7 | 76 | 250 | 0 | 12 | 7.25 | 27 | 539 | 2 |
| 4 | 1.49 | 3.34 | 36.45 | 0.98 | 414.4 | 84 | 300 | 1 | 21.4 | 7.78 | 25 | 628 | 7.76 |
| 5 | 1.4 | 7.51 | 34.17 | 0.67 | 431.6 | 92 | 310 | 1 | 13 | 7.36 | 28 | 654 | 4 |
| 6 | 1 | 3.45 | 35.44 | 0.96 | 353.1 | 64 | 230 | 0 | 16.4 | 7.68 | 27 | 535 | 0.84 |
| 7 | 1.5 | 3.54 | 25.7 | 1.02 | 352.4 | 64 | 230 | 0 | 17.3 | 7.7 | 28 | 534 | 1 |
| 8 | 0.3 | 0.17 | 31.6 | 0.85 | 425 | 100 | 80 | 0 | 0 | 7.73 | 30 | 1022 | 0 |
| 9 | 0.8 | 2.84 | 34.86 | 0.66 | 449 | 125 | 482 | 0 | 0 | 7.62 | 32 | 628 | 5.9 |
| 10 | 0.69 | 9.82 | 26.1 | 0.56 | 448.8 | 92 | 314 | 1 | 16.3 | 7.76 | 28 | 680 | 4.55 |
| 11 | 0.89 | 2.47 | 32.9 | 0.56 | 413.8 | 56 | 292 | 1 | 19.3 | 7.73 | 29 | 627 | 8.1 |
| 12 | 1.5 | 9.77 | 30.06 | 0.57 | 369.6 | 80 | 240 | 0 | 12.4 | 7.36 | 23 | 560 | 0 |
| 13 | 1.74 | 1.59 | 38.6 | 0.9 | 413.8 | 84 | 272 | 1 | 19.8 | 7.79 | 28 | 627 | 8.81 |
| 14 | 1.35 | 3.32 | 34.33 | 0.71 | 341.8 | 56 | 204 | 0 | 14.9 | 7.79 | 22 | 518 | 8 |
| 15 | 1.36 | 3.45 | 35.12 | 0.25 | 534 | 121.7 | 321 | 0 | 22 | 7.7 | 28 | 516 | 19.9 |

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| | | | | | | | | | | | | Table- | 4 (contd.) |
|-----------------------|------|------|-------|------|-------|-------|-------|---|------|---------|----|--------|------------|
| 16 | 1 | 0.97 | 27.8 | 0.45 | 556 | 121.7 | 347.2 | 0 | 16.3 | 7.43 | 32 | 1027 | 0.25 |
| 17 | 1.86 | 1.5 | 34.87 | 0.54 | 412.9 | 79 | 320 | 0 | 11 | 7.34 | 27 | 760 | 4.35 |
| 18 | 1 | 1.07 | 33.14 | 0.22 | 505 | 138 | 105 | 0 | 0 | 7.26 | 34 | 449 | 6.2 |
| 19 | 1.86 | 5.78 | 37.78 | 0.68 | 560 | 109 | 340 | 0 | 11 | 7.56 | 28 | 780 | 6.98 |
| 20 | 0.98 | 5.37 | 38.35 | 0.56 | 389 | 125 | 270 | 0 | 0 | 7.68 | 28 | 690 | 5.76 |
| Egyptian standards | 0.5 | 45 | 250 | 0.3 | 1000 | 250 | 500 | - | - | 6.5–8.5 | - | - | 0–1 |

| Table 5 | . Average cou | nts of total bacteria | al counts, bacterial i | ndicators, total fun | gi and yeast in gro | undwater sample | es ⁶ | |
|--------------------|---------------|-----------------------|------------------------|----------------------|---------------------|-----------------|-----------------|--|
| Sample | TBC (| cfu/ml) | тс | FC | FS | FF | Total yeasts | |
| | At 37°C | At 37°C | (cfu/100 ml) | (cfu/100 ml) | (cfu/100 ml) | (cfu/100 ml) | (cfu/100 ml) | |
| 1 | 52 | 61 | 6 | 3 | 6 | 22 | 31 | |
| 2 | 70 | 79 | 9 | 1 | 5 | 9 | 60 | |
| 3 | 148 | 167 | 46 | 25 | 12 | 97 | 15 | |
| 4 | 90 | 112 | 12 | 7 | 4 | 56 | 20 | |
| 5 | 118 | 125 | 32 | 16 | 8 | 78 | 19 | |
| 6 | 94 | 108 | 17 | 8 | 6 | 64 | 0 | |
| 7 | 115 | 119 | 21 | 10 | 1 | 89 | 84 | |
| 8 | 50 | 57 | 0 | 0 | 0 | 7 | 3 | |
| 9 | 16 | 25 | 0 | 0 | 0 | 4 | 12 | |
| 10 | 63 | 72 | 7 | 2 | 6 | 8 | 19 | |
| 11 | 152 | 170 | 29 | 12 | 4 | 96 | 180 | |
| 12 | 58 | 72 | 14 | 6 | 1 | 77 | 37 | |
| 13 | 60 | 69 | 4 | 1 | 2 | 9 | 130 | |
| 14 | 64 | 80 | 18 | 4 | 3 | 29 | 211 | |
| 15 | 84 | 120 | 15 | 5 | 4 | 20 | 75 | |
| 16 | 103 | 135 | 18 | 9 | 1 | 47 | 5 | |
| 17 | 126 | 148 | 37 | 13 | 6 | 89 | 92 | |
| 18 | 60 | 81 | 10 | 23 | 0 | 119 | 56 | |
| 19 | 75 | 94 | 20 | 19 | 5 | 77 | 0 | |
| 20 | 70 | 85 | 40 | 21 | 3 | 97 | 16 | |
| Egyptian standards | ≤ 50 | ≤ 50 | ≤ 2 | ≤ 1 | ≤ 1 | | | |

ture of filamentous fungi along with fungal spores³². However, mycotoxin producing species are filamentous *ascomycetes, basidiomycetes* and *deuteromycetes* with *Pencillium, Aspergillus and Fusarium* are the most mycotoxin producing fungi³³. Chemical structure of mycotoxins vary significantly, basically contain low mass organic compounds. In other words, mycotoxins are minute and stable molecules, even, problematic to eliminate it which enters into the food chain³⁴. Even though, mycotoxins are also ingested through food or water infested with poisonous fungi³⁵ or ingested as mycotoxins secreted by fungi without eating the fungus itself. Then mycotoxins cause tissue destructions, thrombosis, infarction, and other manifestations of mycosis³⁶ (Table 11).

Water treatment procedures and inactivation of fungi

Different methods are accounted for inorder to inactive the fungi and their mycotoxins from water, especially from drinking water.

Coagulation and flocculation: Sedimentation and flotation process remove many microorganisms and fungi as they are trapped within the particles and settled down at the

| | Table 6. Total fungal count and relative density isolated from groundwater samples ⁶ | | | | | | | |
|----------------------------|---|----------|--|--|--|--|--|--|
| Fungal sp. | Total count cfu/100 ml | R.D.*(%) | | | | | | |
| Filamentous fungi | | | | | | | | |
| Achlya debaryana | 30 | 1.5 | | | | | | |
| Asperigillus flavus | 271 | 13.4 | | | | | | |
| Asperigillus fumigatus | 78 | 3.8 | | | | | | |
| Asperigillus niger | 166 | 8.2 | | | | | | |
| Asperigillus terreus | 89 | 4.4 | | | | | | |
| Rhizopus nigricans | 38 | 1.9 | | | | | | |
| Fusarium oxysporum | 68 | 3.4 | | | | | | |
| Mucor sativas | 54 | 2.7 | | | | | | |
| Paecilomyces varioti | 119 | 5.9 | | | | | | |
| Penicillium notatum | 65 | 3.2 | | | | | | |
| Penicillium egyptiaccum | 116 | 5.7 | | | | | | |
| Total yeasts | | | | | | | | |
| Candida albicans | 230 | 11.4 | | | | | | |
| Candida glabrata | 200 | 9.8 | | | | | | |
| Saccharomyces cerevisiae | 500 | 24.7 | | | | | | |
| Total fungal count | 2024 | 100 | | | | | | |
| *R.D.: Relative Diversity. | | | | | | | | |

 Table 7. Correlation between filamentous fungi and yeast, and bacterial indicators⁶

| | n | | Correlation | | | | |
|-------------------|----|-----------|-------------|--------------|--|--|--|
| | | Total | Fecal | Fecal | | | |
| | | coliforms | coliforms | streptococci | | | |
| Filamentous fungi | 20 | 0.727 | 0.885 | 0.181 | | | |
| Total yeasts | 20 | 0.075 | -0.154 | -128 | | | |

sedimentation tank, from which it is disposed of with the sludge³⁷.

Filtration methods: One of the rapids and filterismostly used method, do not have more retention time to remove all microorganisms from water which are hidden in the particles³⁸. Fungi grow attached to a substrate and colonise filters in water treatment plants giving them an opportunity to resist water treatment³⁹. However, after filtration, the final and most trusted process for annihilating pathogenic microorganism is disinfection⁴⁰.

Disinfection: The use of disinfection in water treatment as a public health measure has shown a major decline in people contracting water-related diseases from drinking wa-

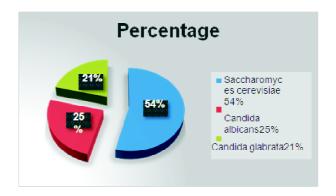


Fig. 2. Relative diversity of total yeasts species in groundwater samples⁶.

| Table 8. Correlation between filamentous fungi and | veasts, and physiochemical parameters ⁶ |
|--|--|
| | youoto, and phyotoononnou paramotoro |

| | | Correlation (n = 20) | | | | | | | | | | | |
|-------------------|--------|----------------------|--------|--------|--------|--------|-------|---------|--------|----------------|--------|-----------|--------|
| | TDS | chlor. | BOD | COD | Temp. | EC | Amm. | Nitrate | Sulph. | Total hardness | Iron | Turbidity | pН |
| Filamentous fungi | -0.081 | -0.087 | -0.082 | -0.098 | 0.146 | -0.231 | 0.161 | -0.091 | -0.002 | -0.237 | -0.153 | -0.152 | -0.366 |
| Total yeasts | -0.295 | -0.517 | 0.2 | 0.367 | -0.303 | -0.354 | 0.218 | -0.222 | 0.106 | -0.141 | -0.017 | -0.4 | 0.298 |

| | Table 9. Isol | ation of fungi from the surface | e water of Veml | panadu wetland agro ecosystem | 24 |
|---------|-------------------|---------------------------------|-----------------|-------------------------------|-------------------------|
| SI. No. | Fungi isolated | Frequency of occurrence | SI. No. | Fungi isolated | Frequency of occurrence |
| | | (%) | | | (%) |
| 1. | A. corymbifera | 5 | 13. | Fusarium sp. | 2.5 |
| 2. | Aletrnaria sp. | 5 | 14. | F. chlamydosporum | 2.5 |
| 3. | A. candidus | 2.5 | 15. | Mucor sp. | 5 |
| 4. | A. flavus | 22.5 | 16. | M. sterilia | 5 |
| 5. | A. fumigatus | 10 | 17. | M. sterilia (demateaceous) | 5 |
| 6. | A. nidulans | 10 | 18. | Paecilomyces sp. | 22.5 |
| 7. | A. niger | 50 | 19. | P. lilacinus | 2.5 |
| 8. | Chrysosporium sp. | 17.5 | 20. | Penicillium sp. | 40 |

| | | | | | Table-9 (contd.) |
|-----|------------------|-----|-----|-----------------|------------------|
| 9. | C. bertholletiae | 2.5 | 21. | P. purpurogenum | 2.5 |
| 10. | Curvularia sp. | 2.5 | 22. | P. verrucosum | 2.5 |
| 11. | C. geniculate | 5 | 23. | S. brevicaulis | 2.5 |
| 12. | C. lunata | 2.5 | 24. | Trichoderma sp. | 2.5 |

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ter⁴¹. Fungi and bacteria have a capability to live longer in verse situation in order to preserve their lives, while the fac-

| Table 10. Quarterly isolated fungi of Vembanadu wetland agro | | | | | | dermination ⁺² . F higher than 1 N | |
|--|--------|----------|----------------|--------|--------|--|-----------------------|
| | 0 ! | ecosyste | em²⁺ | 0 | | | |
| : | Sample | CFU/ | | Sample | CFU/ | Table 11. Fung | gal |
| - : , , | No. | 100 ml | | No. | 100 ml | · | - |
| First quarter | 1 | 3 | Fourth quarter | | 2 | Mycotoxin | G |
| (January-March) | 2 | 6 | (October- | 32 | 8 | Aflatoxins | A |
| | 3 | 8 | December) | 33 | 14 | | |
| | 4 | 12 | | 34 | 0 | | |
| | 5 | 1 | | 35 | 0 | | |
| | 6 | 4 | | 36 | 6 | Fumonisins | F |
| | 7 | 8 | | 37 | 24 | Citrinin | |
| | 8 | 11 | | 38 | 16 | | |
| | 9 | 2 | | 39 | 3 | Ochratoxin A | |
| o | 10 | 1 | | 40 | 9 | | |
| Second quarter | 11 | 30 | | | | Patulin | |
| (April-June) | 12 | 5 | | | | | |
| | 13 | 9 | | | | | A ai s Fi Ti |
| | 14 | 2 | | | | | |
| | 15 | 4 | | | | Sterigmatocystir | ۱A |
| | 16 | 9 | | | | Cyclopiazonic | |
| | 17 | 3 | | | | acid | |
| | 18 | 1 | | | | Trichothecenes | |
| | 19 | 11 | | | | | |
| | 20 | 15 | | | | Deoxynivalenol | |
| Third quarter | 21 | 6 | | | | 2007.9 | |
| (July-September) | | 3 | | | | | |
| | 23 | 12 | | | | T-2 Toxin | F |
| | 24 | 9 | | | | Zearalenone | F |
| | 25 | 3 | | | | Ergot alkaloids | С |
| | 26 | 5 | | | | Penicillic acid | Ρ |
| | 27 | 16 | | | | Tenuazonicacid, | A |
| | 28 | 10 | | | | alternariol, | |
| | 29 | 56 | | | | alternuene, | |
| | 30 | 19 | | | | altertoxin-1 | |
| | 38 | 16 | | | | Ergotamine | С |
| | 39 | 3 | | | | | _ |
| | 40 | 9 | | | | Rhizonin | R |

tors are favourable again, they return to their normal state, recovering the metabolic activity and the generating spore germination⁴². However, the turbidity of the water is getting higher than 1 NTU usually with organic particles. Particles

| | effect | ucing fungal genera and health 22 |
|------------------|--|--|
| Mycotoxin | Genera | Impact on Human Health |
| Aflatoxins | Aspergillus sp. | Human carcinogenic and other are hepatotoxic, aflatoxicosis, as pergillosis, nephropathy, terato genic effect, susceptibility to HIN TB etc. |
| Fumonisins | Fusarium sp. | Oesophageal cancer |
| Citrinin | Aspergillus sp., Penicillium sp. | Nephrotoxic, teratogenic etc. |
| Ochratoxin A | Aspergillus sp. and Penicillium sp. | Hepatotoxic, carcinogenic etc. |
| Patulin | Aspergillus sp., | Toxication on Immune system, |
| | Penicillium sp., | cytotoxic, teratogenic, carcino- |
| | Paecilomyces sp. | genic etc. |
| Sterigmatocystin | Aspergillus sp. | Carcinogenic |
| Cyclopiazonic | Aspergillus sp. | Convulsions |
| acid | and Penicillium sp. | |
| Trichothecenes | Fusarium sp., | Toxic aleukia |
| | Trichoderma sp. | |
| Deoxynivalenol | <i>Fusarium</i> sp. | Anorexia, vomiting, abdomin pain, diarrhoea, giddiness, co vulsion etc. |
| T-2 Toxin | <i>Fusarium</i> sp. | Alimentary toxic aleukia |
| Zearalenone | Fusarium sp. | Carcinogenic |
| Ergot alkaloids | Cladosporium sp. | Ergotism in human body |
| Penicillic acid | Penicillium sp. | Carcinogenic |
| Tenuazonicacid, | Alternia sp. | Inhalation allergy problems an |
| alternariol, | | mycotoxicoses |
| alternuene, | | |
| altertoxin-1 | | |
| Ergotamine | Claviceps purpurea | Neurotoxins (Effect on nervor system) |
| Rhizonin | Rhizopus sp. | Hepatotoxic |

interfere with disinfection as microorganisms are absorbed onto the surface of the particle⁴³.

Ozone: Ozone inactive fungi species by causing irreversible cellular damage⁴⁴. But few isolated species have enough resistant agains to zone activation like *Trichodermaviride* and *Penicillium spinulosum*³⁹.

Free chlorine: Chlorine is a frequently used form of disinfection in water treatment plants in many countries. The chlorine is extensively used as a strong oxidant diminishing harmful microorganisms from water^{45,46,47}.

Conclusion

Hence, it is vast to explain that above-mentioned water samples are not always suitable for drinking purposes as because it is sometimes vulnerable to health. Consequently, it is clearly seen from the tables that different sources of water examined in order to recuperate with the solution. In addition to this, various mechanisms are taken into consideration but not always worthy of treated drinking water. Even though, isolated genera such as Cladosporium, Phoma, Alternia, Aspergillus, Penicillium, Exophila, Fusarium, Acremonium, Exophaila and Phialophora, etc. have an ability to resist against disinfection⁴⁸. So, the documented pathogenic fungi from treated drinking water can no longer be ignored as water contaminants. While many countries are not still aware of the occurrence of fungi and their mycotoxins in treated drinking water supply network²². Monitoring and keeping the amount of fungi under surveillance after water treatment and in the distribution system is fundamental in guiding against harm to human health and also need to develop the aesthetic quality of water like taste and odour. Apart from this, the routine examination can be also challenging because cautious and experienced personnel are needed, though, this should not be underestimated further because fungi influence affecting diversely in waterbodies.

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