

Application of Artificial Neural Network and Particle Swarm Optimization for modeling and optimization of CO₂ sequestration using microalgae

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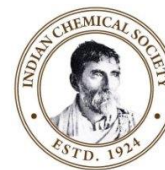
Abstract: In the present study, *Chlorococcum* sp., a green alga – collected from coke-oven effluent treatment plant, was used for the sequestration of carbon dioxide at diverse initial CO₂ concentrations (5-30%), pH of the medium (7-11) and inoculum sizes (5-12.5%), respectively. The test strain showed sustainability in growth upto 30% CO₂. However, the maximum CO₂ sequestration was found to be 79.08±0.89% at 5% CO₂, pH 9 and 12.5% inoculum size for the test strain. Since phenomenological modeling is difficult to build in such systems, artificial neural network modeling technique was tried in this work to predict the CO₂ removal at dissimilar pH of the medium, diverse CO₂ concentrations and at various inoculum sizes for the test strain. Various algorithm of artificial neural network (ANN) model and lot of activation functions were tried to arrive at best ANN model for this experimental data. Developed ANN model performance was found very efficient. Optimization by particle swarm optimization (PSO) algorithm is carried out to determine the optimum value of input parameters to sequester maximum CO₂. With PSO algorithm the optimum input parameters are obtained to achieve 99% removal of CO₂. As far as known the studies on application of Artificial Neural Network and Particle Swarm Optimization for modeling and optimization of CO₂ sequestration using microalgae are very rare and here lies the novelty of the present work.

Keywords: CO₂ sequestration, Green alga, Macromolecules, FAME analysis, ANN

Introduction

CO₂ is a considerable contributor to the global warming and climate change¹. Coal

fired power plants are one of the key sources to atmospheric CO₂². Flue gas releasing from power plants has a temperature of 120°C and CO₂ in the range of 10-15%³. Several



techniques have been engaged so far for biofixation of CO₂, among them biological fixation using microalgae/cyanobacteria has been found as one of the efficient move towards CO₂ sequestration⁴. Cultivation of algae has been exploited as a supplementary move in treatment of flue gas, towards the lessening of CO₂ levels in the exhaust flue gas⁵. Microalgae have the advantages in biofixation of CO₂ over terrestrial plants such as CO₂ fixation in aqueous algal suspension can be achieved in a precise manner and the rate of fixation is considerably higher⁶. The microalgal biomass assemble considerable amounts of lipids, proteins and carbohydrates and some expensive compounds including vitamins and pigments, which can be utilized as an active constituent in pharmacy, feed supplements and food preservatives or in the biofuels production⁵. Further, though a considerable number of research works was done on bio-fixation of CO₂ using microalgae, a comprehensive study encompassing detailed experimentation, data driven artificial intelligence based modeling and optimization of input parameters for maximization of CO₂ removal is very small.

Since application of artificial neural network (ANN) and particle swarm optimization technique in biological system is very sparse, in the present article, both of these techniques have been used to model and optimize the biofixation of CO₂ using a green alga of *Chlorococcum* sp. Parametric study has been done to see the effect of different parameters such pH, initial concentration of CO₂, etc. The data have been used in ANN and finally, PSO is used to get optimum values of parameters for maximum biofixation of CO₂. The studies on application of ANN and PSO for modeling and optimization of CO₂ sequestration using microalgae are very few and here lies the novelty of the present work.

Results and discussion

Growth and lethal dose study for the test strain

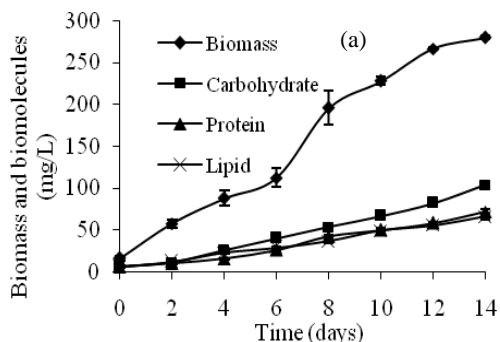


Fig. 1a. Growth of biomass and biomolecules of *Chlorococcum* sp. in BG-11 medium

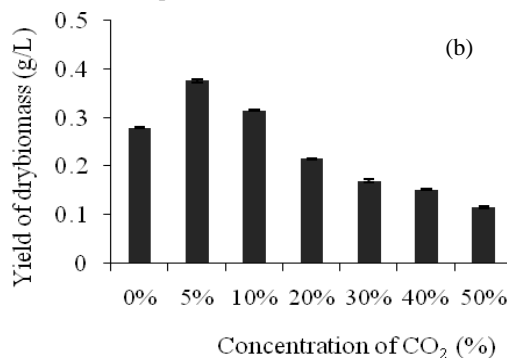


Fig. 1b. Lethal dose analysis of *Chlorococcum* sp. in BG-11 medium

Growth of the isolated green algal strain has been studied in BG-11 medium in terms of biomass yield and thereafter, for the production of lipid, protein and carbohydrate (Fig. 1a). The concentrations of biomass, lipid, protein and carbohydrate have been increased 4.9, 5.4, 7.4 and 9 fold with respect to those concentrations on day 2 for *Chlorococcum* sp. One of the probable reasons for the proliferation may be due to the presence of high concentration of nitrogen (nitrate) and phosphorous in the BG-11 medium⁷. Researchers reported that phosphorous (P) is one of the crucial nutrients in proliferation and metabolism of microalgae⁸. *Chlorococcum* sp. has shown growth upto 50% CO₂ concentration, however, the concentration of biomass is higher (0.315±0.0014 g/L) upto 10% CO₂ than that of the control (0.279±0.002

g/L) (Fig. 1b). The biomass concentration decreases (0.215 ± 0.001 g/L) at and beyond 20% CO₂. The biomass concentration at 5 and 10% CO₂ is 1.34 and 1.13 times higher than that of control. Anjos et al.⁶ reported the biomass growth inhibition at higher CO₂ concentrations.

Optimization of input parameters during CO₂ sequestration

Effect of pH on sequestration of CO₂

Results reveal that with increase in pH from 7 to 9, CO₂ sequestration increases. The sequestration of CO₂ has been found to be $45.79 \pm 0.89\%$, $73.28 \pm 1.34\%$ and $50.81 \pm 0.45\%$ for *Chlorococcum* sp. (green alga) at 5% CO₂, 10% inoculum size at pH 7, 9 and 11, respectively, after 14 days of incubation (Fig. 2a).

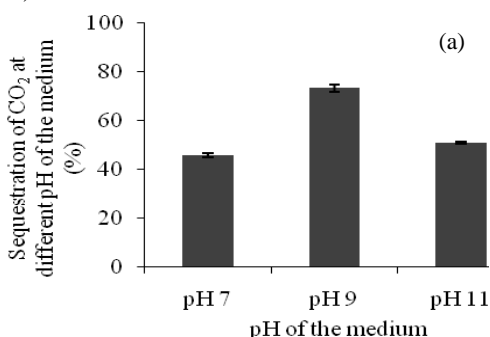


Fig. 2a. Sequestration of CO₂ using *Chlorococcum* sp. at different initial pH of the medium

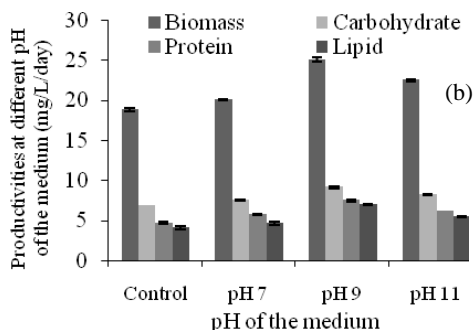


Fig. 2b. Productivities of biomass and biomolecules of *Chlorococcum* sp. at different initial pH of the medium

The maximum productivities of biomass and macromolecules such as lipid,

protein and carbohydrate during optimization of input parameters have been found as 25.07 ± 0.25 , 6.99 ± 0.12 , 7.5 ± 0.15 and 9.16 ± 0.12 mg/L/day for green algae at pH 9 and 10% inoculum size (Fig. 2b). With further raise in pH to 11 the productivities are decreased. It has also been observed that the productivities obtained at dissimilar pH conditions supplemented with CO₂ are found to be maximum than that of control. pH of the medium plays a significant role in growth of the microalgae species, as it has significant effect on the activity of diverse enzymes.

Effect of CO₂ concentration

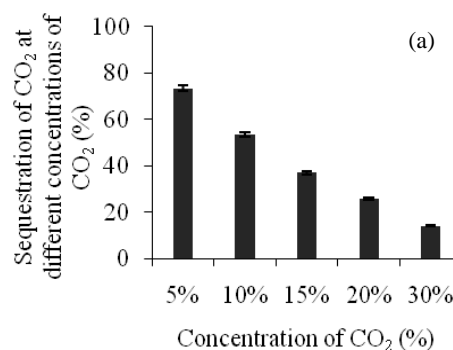


Fig. 3a. Sequestration of CO₂ using *Chlorococcum* sp. at different CO₂ concentrations

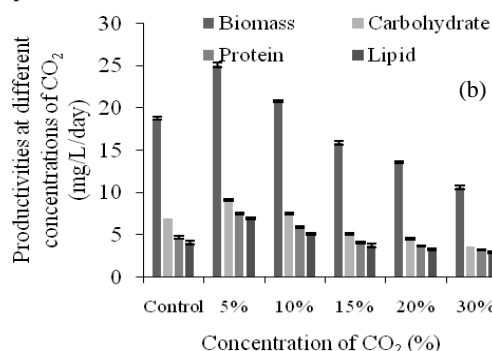


Fig. 3b. Productivities of biomass and biomolecules of *Chlorococcum* sp. at different CO₂ concentrations

The maximum sequestration of CO₂ has been found to be $73.27 \pm 1.34\%$, $53.32 \pm 0.89\%$, $36.87 \pm 0.60\%$, $25.71 \pm 0.34\%$ and $14.01 \pm 0.17\%$ for *Chlorococcum* sp. for the initial CO₂ concentrations of 5%, 10%, 15%,



20% and 30%, respectively, after 14 days under laboratory conditions (Fig. 3a). All other parameters have been maintained constant at pH 9 and 10% inoculum size. It is obvious that higher sequestrations are expected at lower concentrations of CO₂ for the test strain.

The productivities of biomass and macromolecules at dissimilar concentrations of CO₂ have been found to be higher than control upto 10% CO₂ for the green algae (Fig. 3b). The maximum productivities of biomass (25.07±0.25 mg/L/day), lipid (6.99±0.12 mg/L/day), protein (7.5±0.15 mg/L/day) and carbohydrate (9.16±0.12 mg/L/day) have been found at 5% CO₂, 10% inoculum size and pH 9 and it is comparable with the green alga *Chlorococcum littorale*⁹. Also, the productivities of macromolecules at 15, 20 and 30% CO₂ are less than that of control. Though the sequestration of CO₂ is maximum at 5% CO₂, the increase in productivities of biomass, lipid, protein and carbohydrate are 2.06, 2.01, 1.47 and 1.49 fold at 10% CO₂. Since the amount of carbonate present was less (1.3 g/L) at 5% CO₂, lower values of productivities can be expected. Lam et al.⁸ stated that the growth rate of *Chlorella* sp. was inhibited above 2% (v/v) CO₂. Researchers reported that the microalgal cultivation above 5% (v/v) CO₂ is considered as deadly to the growth of microalgae¹⁰.

Effect of inoculum size

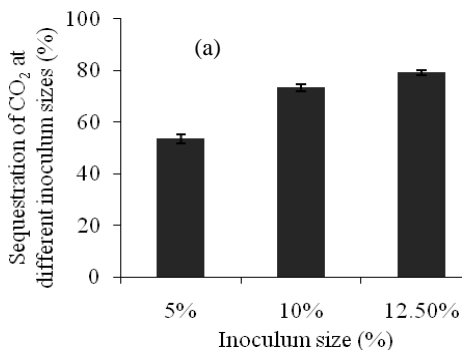


Fig. 4a. Sequestration of CO₂ using *Chlorococcum* sp. at different inoculum sizes

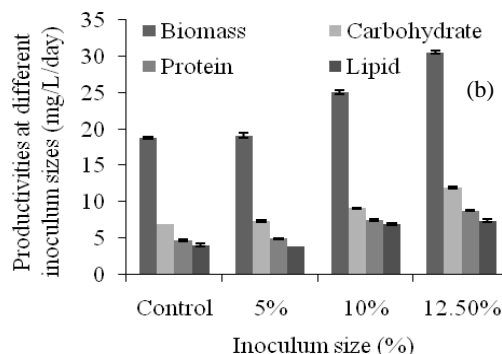


Fig. 4b. Productivities of biomass and biomolecules of *Chlorococcum* sp. at different inoculum sizes

The maximum CO₂ sequestration has been observed as 53.52±1.78%, 73.27±1.34% and 79.08±0.89% for the green alga when the inoculum sizes are at 5%, 10% and 12.5%, respectively (Fig. 4a). Other parameters have been set as pH 9, 5% CO₂. It is obvious that with raise in inoculum size, the number of microalgal cells also increases which eventually leads to the high sequestration of CO₂. Therefore, the productivities of biomass and macromolecules are maximum when the inoculum size is maximum (12.5%). The utmost productivities of biomass (30.54±0.20 mg/L/day), lipid (7.4±0.18 mg/L/day), protein (8.84±0.05 mg/L/day) and carbohydrate (11.95±0.10 mg/L/day) have been found at 12.5% inoculum size, pH 9 and 5% CO₂ for the test strain (Fig. 4b). These productivities are higher than the productivities of *Chlorococcum* sp. when grown in CHU-10 medium¹¹. Researchers reported that the sequestration of CO₂, biomass and macromolecules concentration increased when the inoculum size was varied from 5 to 12.5%¹². Though the sequestration of CO₂ and productivities were found to be maximum at 12.5% inoculum size, effective inoculum size was selected as 10%, since the productivities at 10% and 12.5% inoculum size were comparatively same.

Characterization of the test strain

FTIR study of *Chlorococcum* sp. before and after CO₂ mitigation was performed (Figure not shown). The functional groups at 1056, 1385, 1543, 1648, 2933 and 3408 cm⁻¹ for the green algae when grown in BG-11 medium are responsible for C-O, C-H (bend), N-O, C=O, C-H (stretch) and N-H (amide) bonds. There is no much change in the functional groups of the biomass of green algae after CO₂ uptake.

Time variation of CO₂ sequestration using microalgae

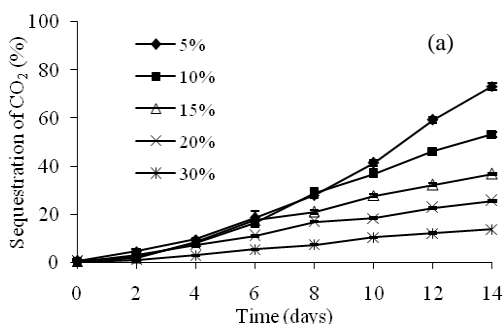


Fig. 5a Sequestration of CO₂ using *Chlorococcum* sp. at different CO₂ concentrations

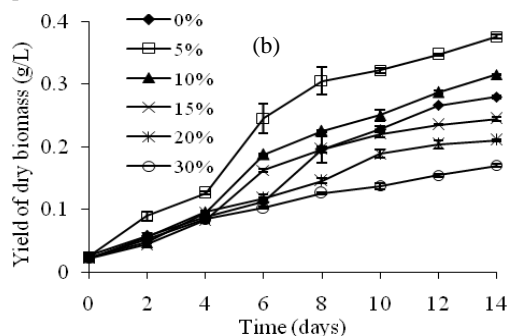


Fig. 5b. Yield of dry biomass of *Chlorococcum* sp. at different CO₂ concentrations

Time variation study of CO₂ sequestration has been done at different CO₂ concentration (5-30%) while, pH and inoculum size have been set as 9 and 10% for 14 days. The CO₂ sequestration decreases from 73.27±1.34 to 14.01±0.17% for change of CO₂ concentration from 5 to 30% (Fig. 5a). Since the amount of carbonate present was less (1.3

g/L) at 5% CO₂, higher sequestration could be expected.

The concentration of biomass has been increased from 0.28±0.002 to 0.315±0.001 g/L with variation in concentration of CO₂ from 0 to 10% (Fig. 5b). The biomass concentration has been decreased to 0.17±0.003 g/L with further increase in CO₂ concentration to 30%. However, the maximum concentration of biomass (0.376±0.0014 g/L) has been observed at 5% CO₂. The reduction in biomass growth has been observed for the test strain supplemented at elevated CO₂ concentrations (15%), may be allied due to some changes in the photosynthetic characteristics of the microalgal strain used. Researchers reported that the CO₂ sequestration and microalgal growth was inhibited at high concentrations of CO₂⁵. When microalgae proliferated at high concentrations of CO₂ (15% or more), several microalgae showed lower uptake of CO₂, higher photosynthetic warmth to O₂ and lesser activity of CA, the enzyme which is responsible for photosynthetic exploitation of inorganic carbon source¹³. As per previous reports the maximum biomass production of *Chlorella* sp. was found to be 0.099±0.001 g/L when grown at 15% CO₂ under high density inoculum¹⁴. However, in the present study, the green alga has shown 0.25±0.003 g/L of biomass at 15% CO₂. The CO₂ bio-fixation and biomass production vary markedly depending on the uniqueness of microalgal species and cultivation conditions¹⁵.

Evaluation of macromolecules content

For the present strain, the lipid concentration increases from 65.71±3.3 to 75.7±1.7 mg/L when the CO₂ concentration varies from 0 to 10% and decreases to 45.72±1.6 mg/L with further increase in CO₂ concentration to 30%. However, the maximum concentration of lipid has been found as



104.28± mg/L at 5% CO₂, pH 9 and 10% inoculum size (Fig. 5c).

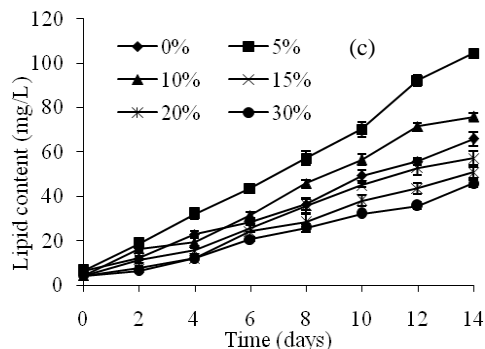


Fig. 5c. Lipid content of *Chlorococcum* sp. at different CO₂ concentrations

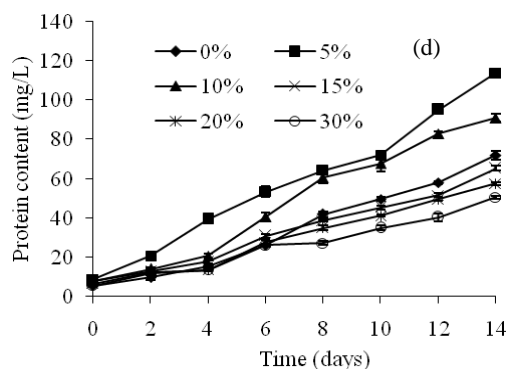


Fig. 5d. Protein content of *Chlorococcum* sp. at different CO₂ concentrations

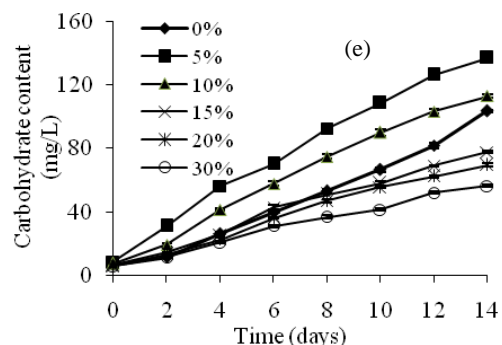


Fig. 5e. Carbohydrate content of *Chlorococcum* sp. at different CO₂ concentrations

The carbohydrate and protein concentrations follow the same trend as that of biomass and lipid content under identical condition. The protein content increases from 71.87±2.2 to 113.75±2.2 mg/L with increase in CO₂ concentration from 0 to 5% for green alga

(Fig. 5d). The concentration of protein decreases to 50.63±0.72 with further increase in CO₂ to 30% for the test strain. The maximum concentration of carbohydrate has been found as 137±1.7 mg/L at 5% CO₂ for green alga (Fig. 5e) after 14 days. With further increase in CO₂ concentration 15 to 30%, the carbohydrate content decreases.

FAME analysis

The fatty acid contents suitable for the production of biodiesel (C16:0) have been found as 43.26% and 46.64% for *Chlorococcum* sp. before and after sequestration of CO₂, respectively (Figures not shown). These amounts have been found to be higher than that of *Scenedesmus* sp. at 15% CO₂¹⁶. The fatty acid contents (C16:0) for *C. vulgaris*, *B. terribilis* and *B. braunii* were found to be 34.82%, 35.46% and 30.39%, respectively, which were lower than that of the present test strain under diverse CO₂ concentrations¹⁷.

Development of a Suitable ANN Model and Optimization using PSO Model

ANN optimization

Typical input-output data has been used in ANN training of CO₂ sequestration for the model performance (Data not shown). The performance of the developed model is as follows; total number of data point: 133, optimum number of node: 15, activation function: Sigmoidal (Hidden layer) and linear (output), training algorithm: Levenberg-Marquardt and prediction accuracy of R^2 (0.999) and very low MSE (1.18). These values suggest the good fitting of experimental data with the ANN model predicted ones. Thus, the suitability of proposed ANN is established.

PSO optimization

Maximum and minimum values of input parameter are fixed as follows: pH of the



medium [11, 7], initial CO₂ concentration [30%, 5%], inoculum size [12.5%, 5%] and time [14 days, 2 days], respectively. The values of four input parameters will be assumed in these ranges and PSO passes these values to ANN trained model to get optimum output.

PSO algorithm keeps on changing until percentage CO₂ removal is maximized¹⁸. Five sets of optimum input parameters are obtained using PSO algorithm to achieve 100% removal (Table 1). The advantage of PSO algorithm lies in the fact that one can get optimum values by performing minimum number of experimentations.

Experimental

Growth and lethal dose study for the test strain

The growth of the test strain was studied in BG-11 medium (NaNO₃: 1.5 g/L, K₂HPO₄: 0.04 g/L, MgSO₄·7H₂O: 0.075 g/L, CaCl₂·2H₂O: 0.036 g/L, Citric Acid: 0.006 g/L, Ferric ammonium citrate: 0.006 g/L, Na₂CO₃: 0.02 g/L, EDTA: 0.001 g/L, Trace metal: 1 mL/L)¹⁹. The test strain was cultivated in different conical (Erlenmeyer) flasks. Finally, the flasks were set aside in algal incubator under laboratory conditions such as temperature: 25°C, light intensity: 2400 Lux, dark/light cycle ratio: 8 h/16 h. The samples were withdrawn at regular time intervals and the biomass was collected from broth by centrifugation at 5000 rpm for 15 min (ELTEK TC8100F, India). The collected microalgal biomass was dried out in an air oven at 60°C for 12 h. The produced biomass was used for the production of lipid, protein and carbohydrate using standard methods¹⁹. The lethal dose analysis was performed to find out the sustainability of the test strain under CO₂ stress condition by growing it in dissimilar CO₂ concentrations in BG-11 medium. The specific CO₂ concentrations were prepared by dissolving the sodium

carbonate salt in BG-11 equivalently using stoichiometric relations¹².

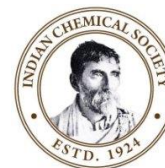
$$\log [\text{HCO}_3^-] = \text{pH} - \log [\text{CO}_2] - 6.30 \quad (1)$$

$$\log [\text{CO}_3^{2-}] = \text{pH} - \log [\text{HCO}_3^-] - 10.09 \quad (2)$$

At pH 8, the values of sodium carbonate concentrations equivalent to 5%, 10%, 20%, 30%, 40% and 50% CO₂ concentrations in the BG-11 medium are 10.9 g/L, 21.8 g/L, 43.6 g/L, 65.5 g/L, 87.3 g/L and 109.2 g/L, respectively. The microalgal strain was grown at different CO₂ concentrations separately. The microalgal growth was investigated for two weeks (14 days), the desired concentrations of CO₂ for the test strain were found based on the biomass growth.

Optimization of input parameters during CO₂ sequestration

Biofixation of CO₂ was done by one factor at a time analysis (OFAT) as described by Upendar et al. (2018)²⁰. Initially, pH was varied from 7-11 and all other parameters such as initial CO₂ concentration and inoculum size were set invariable at 5% and 10%, respectively. Initial CO₂ concentration was varied later from 5-30% keeping other variables at their corresponding values. Finally, inoculum size was varied from 5-12.5% keeping other variables at their adequate values. The solutions prepared were cultivated with the test strain under laboratory conditions and were grown for two weeks. The cultured samples were drawn after 14 days and the supernatant was collected by centrifugation. The collected supernatant was analysed for carbonate concentration by titration using 0.1 N Hydrochloric acid. All the adequate values of initial CO₂ concentration, inoculum size and pH of the medium were found on the basis of biofixation of CO₂ and biomass growth. The biomass produced after CO₂ biofixation was utilised for the production of macromolecules. The biomass



productivities were calculated using Equation 3:²¹

$$P = \frac{(x_1 - x_0)}{(\theta_1 - \theta_0)} \quad (3)$$

Where, θ_0 = 0th day (days), θ_1 = beginning of stationary phase (days), x_0 = Biomass and biomolecules concentration on 0th day (mg/L), x_1 = Biomass and biomolecules concentration after log phase (mg/L).

Characterization of cellular biomass

Fourier Transform Infrared (FTIR) study of cellular biomass was done for the test strain to get the availability of organic groups on the surface of cellular biomass before and after biofixation of CO₂. For the FTIR study, two types of samples were prepared. At first, test strain was grown in BG-11 with 10% inoculum, pH 9 and 25°C temperature (control), subsequently the test strain was grown in BG-11 medium supplemented with CO₂ (5%). The samples were grown for two weeks in incubator under laboratory conditions. After two weeks the biomass was collected using centrifugation and dried in hot air oven and thereafter used for the FTIR examination.

Time variation of CO₂ biofixation using test strain

Time variation of biofixation of CO₂ was studied at various carbon dioxide concentrations (5-30%). All other variables like pH, inoculum size and temperature were set at their most effective conditions i.e., the conditions at which the sequestration of CO₂ and concentration of biomass and biomolecules were found to be maximum. The solutions were inoculated using the test strain and were placed in incubator under laboratory condition for 14 days. The biomass generated was utilised for the production of macromolecules using standard methods as

mentioned in 'Growth and lethal dose study for the test strain' subsection.

FAME analysis

Fatty acid methyl esters (FAMES) are the molecules that are principally present in biodiesel. FAME analysis reveals that the fraction of lipid content of the test strain can directly be used as biodiesel production. Total lipid content was extracted from dry biomass before and after biofixation of CO₂. The total lipid extracted and its fractions such as phospholipids (PL), glycolipids (GL) and neutral lipids (NL) were transesterified into FAMES as described by Mistry et al.¹¹.

Development of a suitable ANN model and optimization using PSO model

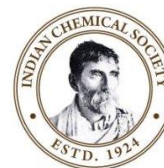
Development of ANN model

Experiments were carried out to validate CO₂ sequestration using test strain. The following four parameters were considered which have large effect on biofixation of CO₂ using algal strains: (i) time of contact, (ii) initial CO₂ concentration, (iii) pH, (iv) inoculum size. Percentage of CO₂ removal is taken as target. For development of ANN model, the algorithm as discussed by Upendar et al.¹⁸ was followed. The topology consists of four input nodes and one output node. Best ANN algorithm was selected during building time.

The aptness of ANN model is examined by MSE and coefficient of determination (R^2). Less than one hour was required for calculation of this large number of ANN models using intel i-5 processor.

Development of a suitable PSO model

Once an accurate and reliable ANN model was built up, the next optimization algorithm is tried to optimize the input parameters namely time, pH, initial concentration, inoculum size and so that the percentage removal of CO₂ is maximized.



Since the ANN equations are complicated and comprises of various exponential terms and weights, normal optimization algorithm is difficult to apply on ANN model. For development of PSO algorithm the steps of Upendar et al.¹⁸ were followed to maximize CO₂ sequestration.

Conclusion

CO₂ mitigation using *Chlorococcum* sp. has been found to be a promising option for CO₂ fixation and higher biomass yield. The maximum biofixation of CO₂ has been observed as 79.08±0.89% at 5% CO₂, 12.5% inoculum size and at pH 9, respectively. The strain could grow well upto 30% CO₂. The higher productivities of lipid, protein and carbohydrate indicate the possibility of usage of such strain for able CO₂ biofixation and value-added production. In this study, various ANN model with different algorithms have been evaluated to predict CO₂ sequestration. Very low RMSE and high R² value of the final ANN model indicates that developed model has successfully captured the nonlinear relationship between CO₂ fixation and initial concentration, inoculum size, pH of the medium and time. The PSO algorithm is then applied on ANN model to optimize the input parameters, so that CO₂ removal is maximized. The results of PSO optimization is very encouraging as it shows that more than 99% CO₂ removal is possible by optimizing the initial concentration, inoculum size and pH of the medium. The advantage of PSO optimization is to determine the optimum value of input parameters automatically and thus, it can avoid lots of experimentation. Such ANN modeling and PSO optimization technique can be extended to any other complex system where phenomenology is fully understood.

Conflict of interest

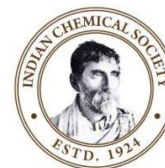
On behalf of all the authors, I declare that there is no conflict of interest.

Acknowledgements

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References

1. N. M. Langely, S. T. L. Harrison, and R. P. van Hille, *Biochem. Eng. J.*, 2012, **68**, 70-75.
2. D. A. Roberts, N. A. Paul, M. I. Bird, and R. de Nys, *J. Environ. Manage.*, 2015, **153**, 25-32.
3. G. Yadav, A. Karemore, S. K. Dash, and R. Sen, *Bioresour. Technol.*, 2015, **191**, 399-406.
4. B. Wang, Y. Li, N. Wu, C. Q. Lan, *Appl. Microbiol. Biotechnol.*, 2008, **79(5)**, 707-718.
5. M. Anjos, B. D. Fernandes, A. A. Vicente, J. A. Teixeira, and G. Dragone, *Bioresour. Technol.*, 2013, **139**, 149-154.
6. P. Chelf, L. M. Brown, and C. E. Wyman, *Biomass & Bioenergy*, 1993, **4(3)**, 175-183.
7. A. Goli, A. Shamiri, A. Talaiekhosani, N. Eshtiaghi, N. Aghamohammadi, and M. K. Aroua, *J. Environ. Manage.*, 2016, **183**, 41-58.
8. M. K. Lam, K. T. Lee, and A. R. Mohamed, *Int. J. Greenh. Gas Cont.*, 2012, **10**, 456-469.
9. S. P. Singh, and P. Singh, *Renew. Sustain. Energ. Rev.* 2014, **38**, 172-179.
10. R. Ramanan, K. Kannan, A. Deshkar, R. Yadav, and T. Chakrabarti,



- Bioresour. Technol.*, 2010, **101(8)**, 2616-2622.
11. A. N. Mistry, G. Upendar, S. Singh, J. Chakrabarty, G. Bandyopadhyay, K. C. Ghanta, and S. Dutta, *Sep. Sci. Technol.*, 2020, **55(2)**, 332-345.
 12. G. Upendar, A. N. Mistry, R. Das, S. G. Thakurata, J. Chakrabarty, K. C. Ghanta, and S. Dutta, *Environ. Prog. Sustain. Energy*, 2018, **37(5)**, 1594-1600.
 13. J-R. Xia, and K-S. Gao, *J. Integr. Plant. Biol.*, 2005, **47**, 668-675.
 14. S. Y. Chiu, C.Y. Kao, C. H. Chen, T. C. Kuan, S. C. Ong, and C. S. Lin, *Bioresour. Technol.*, 2008, **99(9)**, 3389-3396.
 15. U. B. Singh, and A. Ahluwalia, *Mitig. Adapt. Strat. GL.*, 2013, **18(1)**, 73-95.
 16. R. Tripathi, J. Singh, and I. S. Thakur, *Renew. Energ.*, 2015, **74**, 774-781.
 17. I. A. Nascimento, I. T. D. Cabanelas, J. N. dos Santos, M. A. Nascimento, L. Sousa, and G. Sansone, *Algal Res.*, 2015, **8**, 53-60.
 18. G. Upendar, S. Singh, J. Chakrabarty, K. C. Ghanta, Md. Shahnawaz, S. K. Lahiri, and S. Dutta, *Water Environ. J.*, 2020, DOI:10.1111/wej.12646.
 19. K. Anjana, A. Kaushik, B. Kiran, and R. Nisha, *J. Hazard. Mater.*, 2007, **148(1-2)**, 383-386.
 20. G. Upendar, S. Singh, J. Chakrabarty, K. C. Ghanta, S. Dutta, and A. Dutta, *J. Environ. Manage.*, 2018, **218**, 234-244.
 21. D. Tang, W. Han, P. Li, X. Miao, and J. Zhong, *Bioresour. Technol.*, 2011, **102(3)**, 3071-3076.

Table 1. Multiple optimum responses and corresponding operating parameter obtained by Particle Swarm Optimization (PSO) for the removal of CO₂ using *Chlorococcum* sp.

S. No	Time (days)	Initial concentration (%)	Inoculum size (%)	pH	Sequestration of CO ₂ (%)
1	13.99	5.04	12.50	11.00	100.00
2	13.99	5.00	12.50	11.00	100.00
3	14	5.00	12.50	10.99	100.00
4	13.99	5.00	12.50	10.99	100.00
5	14	5.004	12.50	11.00	100.00